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# MITIGATION OF SALT STRESS IN WHEAT PLANT (*Triticum aestivum*) BY PLANT GROWTH PROMOTING RHIZOBACTERIA FOR ACC DEAMINASE

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Wheat (*Triticum aestivum*) is the essential diet of population as it constitutes 60% of the daily diet of common man in Pakistan. Salinity is a barrier towards growing a sustainable food production system and environment management. PGPR (Plant Growth Promoting Rhizobacteria) under salt stressed conditions causes 1-aminocyclopropane-1-carboxylate (ACC) deaminase action which minimizes the intensity of ACC and endogenous ethylene justifying the toxic effects of salt stress on plant growth. The plants inoculated with PGPR having ACC deaminase are relatively more tolerant to salt stress. The study was conducted at National Agriculture Research Centre, Islamabad to examine the consequence of PGPR on wheat crop under saline environment to see that bacterial strains having ACC deaminase had significant effect on wheat growth and ionic concentration. The design was completely randominzed with three repeats. Wheat seeds were inoculated with rhizobacterial strains which were: WPR-51, WPR-61, WM-4, WM- and WPS-8. Salinity (9.68 dS m<sup>-1</sup>) was artificially developed using salts. Growth of wheat plants performed better under saline environment as inoculated with different rhizobial strains due to the production of ethylene under stressed conditions. Reduction in sodium uptake in plants with the inoculation of different rhizobial strains to wheat seed under saline environment is a positive sign to mitigate salt stress biologically.

**KEYWORDS**: Wheat growth, Salinity, Ethylene, Rhizobial strains, Salt tolerance.

# INTRODUCTION

Wheat (*Triticum aestivum*) is the essential diet of population as it constitutes 60% of the daily diet of

common man in Pakistan and average per capita consumption is about 125 kg and occupies a central position in agricultural policies of the government (Government of Pakistan, 2013). Salinity is a barrier towards growing a sustainable food production

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system and environment management throughout the world. Such dilemma has its origin from aquatic, ecological and anthropogenic actions. Soil salinity severely deteriorates soil health (socio-economic) which in turn declines crop productivity. Arid and semi-arid lands worldwide have been seriously affecting the problem of soil. Approximately the increase in saline soils is at the rate of 7% in the world was reported by (Tester and Davenport, 2003). At a global level, the total amount of saline soils is around 15% in arid and semi-arid regions and approximately 40% in irrigated lands (Zahran, 1997). Elevated soil salinity negatively affects the physical and chemical properties of soil, thereby openly affecting the growth and assortment of organisms that live in or on soil such as plants, microbes, protozoa and nematodes. In plants, continuing high soil salinity situation causes ionic and osmotic stress that harmfully affects the functioning of various biochemical processes (Parida and Das, 2005). Under high salinity conditions, plants survive with stress which ultimately restricts expansion of leaves. This shows that cell division and expansion processes are sternly affected firstly besides the closure of stomata (Munns, 2002; Flowers, 2004). Premature ageing of leaves during ionic tress affects photosynthesis process and result in stunted growth (Cramer and Nowak, 1992). More, excessive concentrations of sodium and chloride negatively affect the energy production and physiology of the plants by snooping various enzymes activities (Larcher, with 1980). Significant decrease in productivity of saltsensitive and salt-tolerant crops resulted in salt stress conditions. Mostly the salt stress thresholds of the cereal crops is low e.g. wheat can tolerate salinity up to 6 dSm<sup>-1</sup>, while the salinity threshold for maize is three times less (approximately 2 dSm<sup>-1</sup>) (Saeed et al., 2001). Kotuby-Amacher et al., (2000) exposed that, useful microorganisms can lessen salt stress in maize and wheat crops around 50%. In addition, it has been established that positive microorganism play a significant role in alleviating salt stress in plants, causing in better crop produce. Plant-growth-promoting (PGP) bacteria are a set of microorganisms that colonise the root of plants or free-living organisms that directly or indirectly the growth of plants (Lugtenberg and Kamilova, 2009; Ahmad et al., 2013). In direct growth promotion, they produce some compounds (indole acetic acid, siderophore, HCN, etc.), solubilise minerals and break organic matters for easy uptake by plants and

for their own use. They also fix atmospheric nitrogen produce siderophores that increase the bioavailability of iron and synthesise phytohormones such as cytokinins, auxins and gibberellins which have beneficial roles in various stages of plant growth (Lucy et al., 2004; Gray and Smith, 2005; Richardson and Simpson, 2011). Ultimately, they help in lessening or inhibiting the injurious effects of pathogenic organisms by increasing the host resistance to pathogenic organisms (Bloemberg and Lugtenberg, 2001; van Loon, 2007).

Growth promoting rhizobacteria (PGPR) are considered as useful bacteria in the rhizosphere which are helpful for sustainable agriculture by assisting plant growth and development directly or indirectly (Muhammad et al., 2007). PGPR exert some of these functions by means of specific enzymes, aggravating certain physiological and biochemical changes in plants (Bashan et al., 2004). Hass and Keel, (2003) classified PGPR) based on their activities as biofertilizers (increasing the availability of nutrients to plant), phyto stimulators (plant growth promoting, usually by the production of phytohormones), rhizoremediators (degrading organic pollutants) and bio pesticides (controlling diseases, mainly by the production of antibiotics and antifungal metabolites). Bhattacharyya and Jha, 2012 reported that PGPR are the rhizosphere bacteria that can enhance plant growth by a wide variety of mechanisms like phosphate solubilization, siderophore production, biological nitrogen fixation, rhizosphere engineering, phytohormone production, exhibiting antifungal activity, production of volatile organic compounds (VOCs), induction of systemic resistance. promoting beneficial plant-microbe symbioses and interference with pathogen toxin production.

Ethylene is the plant growth regulating hormone produced in response to water logging salinity and/or drought. PGPR from stressed environment exhibit 1-aminocyclopropane-1-carboxylate (ACC) deaminase activity which reduces the level of ACC and endogenous ethylene mitigating the deleterious effects of stress on overall plant growth. The plants inoculated with PGPR having ACC deaminase are relatively more tolerant to environmental stress (Naveed et al., 2008). Keeping in view of these constrints in saline environment pot culture experiment was conducted to see the response of Plant Growth Promoting Rhizobacteria for ACC-Deaminase activity to induce salt tolerance in wheat (Triticum aestivum) crop and determine the extent

**Table1.** Physiochemical characteristics of soil used in the pot experiment.

Characteristics	Unit	Values
рН	-	7.41
Electrical conductivity	(dS m <sup>-1</sup> )	9.68
Organic Matter	(%)	0.523
Na	ppm	303
K	ppm	47
P (AB-DTPA)	ppm	0.68
Ca+Mg	meq/L	19
Carbonate	meq/L	0.5
Bicarbonate	meq/L	0.18
SAR	meq/L	10.08
Soil type	-	Sandy Loam

and degree to which wheat plant growth is affected with the inoculation of different bacterial strains.

# **MATERIALS AND METHODS**

A pot experiment was carried at National Agriculture Research Centre, Islamabad to see the effect of **PGPR** (Plant arowth promoting bacteria Rhizobacteria) having ACC deaminase on wheat (Cv.NARC-13) growth and ionic concentration during November, 2015 to March, 2016 under saline soil (ECe=9.68 dS m<sup>-1</sup>) as indicated in Table 1. Artifical salinity was developed by adding different salts. Completely randomized design was planned with three replications. Wheat seeds were inoculated with rhizobacterial strains: WPR-51, WPR-61, WM-4, WM-9 and WPS-8. Plant samples were collected to see the effect of different rhizobial strains on the availability of nutrients to plants. Soil samples were analyzed for various physicochemical properties using standard methods (Ryan et al., 2001; Sparks et al, 1996) and soil texture (Kanwar and Chopra 1959). The data obtained were subjected to statistical analysis using the statistical software (Version 8.1) and the mean values were compared using Least significant difference (LSD) multiple range test P: 0.5%.(Steel and Torrie, 1997).

### RESULTS AND DISCUSSION

Plant height significantly affected by the inoculation wheat seeds with different rhizobial strains under artificially developed saline soil i.e. ECe= 9.86 dS m<sup>-1</sup>

(Table 2). The highest plant height (13.97 cm) was gained by inoculating WPR-61 which statistically at par with the results attained with WPS-8 and WM-9 and lowest height in plant (10.07) cm) was observed in control i.e. without inoculation. This indicated that inoculation of wheat seed with rhizobial strains showed better responses in plant height mitigating the adverse effects of saline m<sup>-1</sup>). conditions (ECe=9.86 dS Statistically significant results were attained in plant fresh weight data of wheat plants inoculated with strains under saline environment as indicated in Table 2. Maximum fresh weight (2.06 g plant<sup>-1</sup>) was attained by WPR-51. Remaining strains showed negative results comparing with control saline conditions at ECe=9.86 dS m<sup>-1</sup>. Plant dry weight indicated same results as plant fresh weight in Table 2. Significant findings were obtained in chlorophyll contents as indicated in Table 2 where WPR-51 attained the highest value (38.97%). After that control treatment i.e. without inoculation gained this parameter lowering the values of remaining strains. Many researchers have reported better growth in plants inoculated with bacteria containing ACC-deaminase (Mayak et al., 2004; Shaharoona et al., 2006). Ethylene is a stress hormone and is produced at higher concentration under any kind of stress including salinity. It is very likely that the rhizobacterial strains promoted root and shoot growth by lowering the endogenous inhibitory levels of ethylene in roots because of its high ACC metabolizing ability (Kang et al., 2010). Such promising rhizobial strains could possibly be used under field conditions in saline environment where agriculture is exclusively reliant on saline conditions (Hamayun et al., 2010).

When seeds were inoculated with different strains of bacteria having ACC deaminase effect on wheat plant growth under saline conditions (ECe= 9.86 dS m<sup>-1</sup>). Ionic concentration of P (%) in wheat plants showed significant differences among treatments (Table-3). Uptake of P (%) was more (0.18%) by WPS-8 and control showed the lowest (012%). Uptake of K (%) was the highest (3.38 %) by WPS-8 and lowest was determined in control as well as in WPR- (Table 3). Sodium ionic concentration showed significant results among treatments (Table-3). However, Na (%) was the highest in control and lowest by the wheat seed inoculation by WPR-51. This means that reduction in sodium ions in wheat plants using inoculation with rhizobial strains mitigates the salinity and grows wheat plants in

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Treatments	Plant Height	Plant fresh weight	Plant dry weight	Chlorophyll contents
	(cm plant-1)	(g plant <sup>-1</sup> )	(g plant-1)	(%)
Control	10.07 d	0.98d	0.43 d	38.37b
WPR-51	12.33 b	2.06 a	1.01 a	38.97a
WPR-61	13.97 a	1.75 b	0.89 b	37.97bc
WM-4	11.17 c	1.20 cd	0.70 c	29.43c
WM-9	13.68 a	1.31 c	0.80 bc	29.03cd
WPS-8	13.93a	1.09 d	0.48 d	28.77d
LSD(0.5%)	0.37	0.13	0.11	0.56

**Table 2.** Effect of AC C deaminase on wheat growth under saline conditions.

Values followed by same letter(s) are statistically similar at P=0.05 level of significance.

**Table 3.** Effect of ACC deaminse on the ionic concentration of nutrietns in wheat plants.

Treatments	P (%)	K (%)	Na (%)
Control	0.13 d	2.87c	3.01 a
WPR-51	0.12 d	2.87 c	1.78d
WPR-61	0.17 ab	2.17 d	2.94 a
WM-4	0.15 c	2.93c	1.91 c
WM-9	0.16 b	3.10b	2.08 b
WPS-8	0.18 a	3.38a	2.13 b
LSD(0.5%)	0.01	0.16	0.09

Values followed by same letter(s) are statistically similar at P=0.05 level of significance.

better conditions under saline environment. Singh et al., (2013) reported that judicious use of chemicals along with bio fertilizers and organic resources can be helpful in sustaining the crop productivity and soil health.

# CONCLUSION

Growth of wheat plants performs better under saline environment as inoculated with different rhizobial strains due to the production of ethylene under stressed conditions. Reduction in sodium uptake by the utilization of different rhizobial strains under saline environment is a positive sign to mitigate salt stress biologically. The best rhizobial strains were further evaluated in the farmer fields of salt- affected

lands.

# **REFERENCES**

Ahmad M, Zahir ZA, Nazli F, Akram F, Muhammad A and Khalid M (2013). Effectiveness of halotolerant, auxin producing *Pseudomonas* and Rhizobium strains to improve osmotic stress tolerance in mung bean (*Vigna radiata* L.). Braz J. Microbiol., 44:1341–1348

Bashan Y, Holguin G and de-Bashan LE (2004). Azospirillum-plant relationships: physiological, molecular, agricultural, and environmental advances (1997-2003). Can. J. Microbiol., 58(2):521-577.

Bhattacharyya PN and Jha DK (2012). Plant growth-promoting rhizobacteria (PGPR): emergence in agriculture. World J. Microbial Biotechnol., 28:1327–1350.

Bloemberg GV and Lugtenberg BJJ (2001). Molecular basis of plantgrowth promotion and biocontrol by rhizobacteria. Curr. Opin. Plant Biol., 4:343–350.

Cramer GR and Nowak RS (1992). Supplemental manganese improves the relative growth, net assimilation and photosyntheticrates of salt-stressed barley. Physiol. Plant, 84:600–605.

Flowers TJ (2004). Improving crop salt tolerance. J. Exp. Bot., 55:307–319

Government of Pakistan (2013). Agricultural Statistics of Pakistan.

Gray EJ and Smith DL (2005).Intracellular and extracellular PGPR: commonalities and distinctions in the plant–bacterium signaling

- processes. Soil Biol. Biochem., 37:395–412.
- Hamayun M, Khan SA, Shinwari ZK, Khan AL, Ahmed N and Lee I-J (2010). Effect of polyethylene glycol induced drought stress on physio-hormonal attributes of soybean. Pak. J. Bot., 42(2): 977-986.
- Hass D andKeel C (2003). Regulation of antibiotic production in root-colonizing *Pseudomonas spp.* and relevance for biological control of plant disease. Ann. Rev. Phytopathol., 41: 117-153.
- Kang BG, Kim WT, Yun HS and Chang SC (2010). Use of plant growth-promoting rhizobacteria to control stress responses of plant roots. Plant Biotechnol., 4: 179-183
- Kanwar TS and Chopra SL (1959). Practical Agricultural Chemistry. S. Chand and Co., Delhi.
- Kotuby-Amacher J, Koenig K and Kitchen B (2000). Salinity and Plant Tolerance; Available at https://extension.usu.edu/files/publications/publication/AG-SO-03.pdf
- Larcher W (1980). Physiological Plant Ecology. 2nd totally rev. edition ed. Berlin/New York: Springer-Verlag, :33.
- Lucy M, Reed E and Glick BR (2004). Applications of free living plant growth-promoting rhizobacteria. Antonie Van Leeuwenhoek, 86:1–25
- Lugtenberg BJ, Kamilova F (2009). Plant-growth-promoting rhizobacteria. Ann. Rev. Microbiol., 63:541–556.
- Muhammad S, Muhammad A, Sarfraz H and Ahmad SB (2007). Perspective of plant growth promoting rhizobacteria (PGPR) containing ACC deaminase in stress agriculture. J. Ind. Microbiol. Biotechnol., 34:635-648.
- Munns R (2002). Comparative physiology of salt and water stress. Plant Cell Environ., 25:239–250
- Naveed M, Khalid M, Jones DL, Ahmad R and Zahir ZA (2008). Relative efficacy of Pseudomonas spp., containing ACC-deaminase for improving growth and yield of maize (*Zea mays* L.) in the presence of organic fertilizer. Pak. J. Bot., 40(3): 1243-1251.
- Parida AK and Das AB (2005). Salt tolerance and salinity effect onplants: a review. Ecotoxicol. Environ. Saf., 60:324–349.

- Richardson AE and Simpson RJ (2011). Soil microorganisms mediating phosphorus availability update on microbial phosphorus. Plant Physiol., 156:989–996.
- Ryan J, Estefan G and Rashid A (2001). Soil and Plant Analysis Laboratory Manual. International Center for Agricultural Research in the Dry Areas (ICARDA), Islamabad, Pakistan. p 172.
- Saeed MM, Ashraf M, Asghar MN, Bruen M and Shafique MS (2001). Root Zone Salinity Management Using Fractional Skimming Wells With Pressurized Irrigation. Regional Office for Pakistan, Central Asia and Middle East, Lahore. Int. Water Manag. Inst. (IWMI), :46.
- Shaharoona B, Arshad M and Zahir ZA (2006). Effect of plant growth promoting rhizobacteria containing ACCdeaminase on maize (*Zea mays* L.) growth under axenic conditions and on nodulation in mung bean (*Vignaradiata* L.). Lett. Appl. Microbiol., 42: 155-159.
- Singh NK, Chaudhary FK, Patel DB and Triveni E (2013). Effectiveness of Azotobacter Bio-Inoculate for Wheat Grown Under Dry Land Condition. J. Environ. Biol., 34(5): 927-932.
- Sparks DL, Carski TH, Fendorf SE and Toner CVIV (1996). Kinetic methods and measurements. p. 1275-1307. In D.L. Sparks (ed.) Methods of soil analysis: Chemical methods. Soil Science Society of America, Madison, WI.
- Steel RGD and Torrie JH (1997). Principles and Procedure of Statistics. McGraw Hill Book Co., Inc. Singapore, pp: 173–177.
- Tester M and Davenport R (2003). Na+ tolerance and Na+ transport in higher plants. Ann Bot., 91:503–527.
- van Loon LC (2007). Plant responses to plant growth-promoting rhizobacteria. Eur. J. Plant Pathol., 119: 243–254.
- Zahran HH (1997). Diversity, adaptation and activity of the bacterial flora in saline environments. Biol. Fert. Soils, 25:211–223.